Rapid Communication



Cancer and Ras/Raf/ERK Pathway: The Role of New Regulators in Response to Physical Activity, Exercise, and Training

¹Seyed Morteza Tayebi^{*}, ²Karsten Krüger

¹Department of Exercise Physiology, Faculty of Sport Sciences, Allameh Tabataba'i University, Tehran, Iran. ²Department of Exercise Physiology and Sports Therapy, Institute of Sports Science, Justus-Liebig-Universität Gießen, Gießen, Germany.

Submitted 02 February 2022; Accepted in final form 05 March 2022.

ABSTRACT

Background. Cancer is the second and most common cause of death in developed countries, and epidemiological evidence suggests a similar trend in developing countries. Cancer is currently the cause of twelve percent of all deaths worldwide. The ERK pathway contains important modulators suitable for growth and proliferation, especially tumors. Oxidative stress, growth factors, increased intracellular calcium levels, and stimulation of glutamate receptors activate this pathway. The Sprouty/Spred family acts as definitive negative regulators of the Ras/Raf/ERK signal. **View Points.** The Sprouty/Spred family specifically inhibits ERK activity in response to a wide range of factors such as fibroblast growth factor (FGF), platelet growth factor (PDGF), vascular endothelial growth factor (VEGF), brain-derived neurotrophic factor (BDNF), nerve growth factor (NGF), and glial cell-derived growth factor (GDNF). This family can inhibit MAPK activity and affect the downstream level. Typically, these proteins inhibit growth by modulating RTK signaling and suppressing the MAPK_ERK signaling pathway, suppressing cell proliferation, survival, and migration of endothelial cells and increases vascular permeability. **Objectives.** Research on the effect of exercise at various levels on the Sperouty/Spred family in inactive healthy individuals and patients, especially in cancer situations, has not been performed. So, future research can be directed in this way.

KEYWORDS: MAPK Signaling Pathway, Sprouty, Spred, Tumor, Growth, Proliferation.

INTRODUCTION

Phosphorylation is a simple change that occurs when one or more phosphate groups are added to a protein molecule, such as the glycolysis pathway, which is essential in the metabolism of living organisms. This action is usually based on the amino acid serine or threonine, but in rare cases, it is performed on the amino acid tyrosine and sometimes on lysine. The phosphate entry into a protein enters a negative charge, which causes conformation. This binding is done by enzymes called kinases (1). To date, more than a hundred types of protein kinases are known, but one of the most famous is the large family of Mitogen-Activated Protein Kinases (MAPKs) that control numerous cellular functions. Mitogens are signals outside the cell that send messages about mitosis and cell division induction.

The MAPK signaling pathway has been extensively studied in recent years. This pathway is critical in signal transduction from protein kinases and phosphatases. One of the subunits of MAPK is Extracellular signal–Regulated Kinases (ERKs), the most well-known kinase in this pathway (2, 3).

Threonine and tyrosine residues phosphorylated and activated ERK protein kinase (4). This protein activates many proteins and transcriptional factors for different purposes. ERK activation causes the expression of more than 600 genes (5), and its effect on gene expression transcription is achieved either by activating messaging pathways in the cytoplasm or by demyelinating and moving to the nucleus and phosphorylating transcription factors (6). In the cytosol, the ERK exerts its effects in this way, activating proteins such as p90 ribosomal S6 kinase (P90^{RSK}) that transfers to the nucleus and activates the Serum Response Factor (SRF) transcription factors (7). In the nucleus, it can also activate agents such as ETS domain-containing protein (Elk1), Activating Transcription Factor 2 (ATF2), Activating Protein-1 (AP1), cAMP Responsive Element Binding (CREB), and Myocyte-specific enhancer factor 2C (MEF2C) (8-10).

The activity of this transcription factor is expressed by genes involved in various processes biogenesis such as mitochondrial (11).angiogenesis (12), mitotic cell proliferation (13) and meiosis, and post-mitotic function such as differentiation (14), apoptosis (15) and cyclin D expression; helps regenerate muscle cells and repair muscle damage (16). The MAPK pathway, and especially the ERK, can be activated by many factors. Physical activity can also activate this pathway through various mechanisms such as growth factors secretion and muscle tension, oxidants, and pH reduction (16).

On the other hand, the ERK pathway contains important modulators suitable for growth and proliferation. This pathway is activated by intracellular increases in calcium levels, oxidative stress, growth factors, and stimulation of glutamate receptors (17). The MAPK cascading process initiates cell proliferation responses, leading to Ras (Recovery Activation Signal) activation via agonists that stimulate protein kinase C and growth factors that act on tyrosine kinase receptors. In the cascading process, protein kinase C and Ras activate Raf (Rapidly Accelerated Fibrosarcoma) kinase, activating MAPK/ERK Kinase (MEK). Phosphorylated MEK activates and phosphorylates the ERK, which phosphorylates transcription factors, protein substrates, and other protein kinases that are important in promoting cell proliferation and other cellular responses. Altered activity levels of MAPK factors lead to altered transcription of essential genes in the cell cycle (18).

New Regulators of Ras/Raf/Erk Pathway. The Sprouty/Spred family acts as definitive negative regulators of the Ras/Raf/ERK signal (19-22). Drosophila Sprouty was discovered as an FGF signal antagonist in 1998 (20, 21). In mammals, there are four Sprouty homologs (Sprouty 1-4). Sproutys was later shown to suppress ERK activation induced by various growth factors such as FGF, platelet-derived growth factor, VEGF-A, neurotrophic factor, and GDNF in a particular cell type and a specific growth factor (23).

Many factors disrupt Ras/Raf/ERK pathway regulation in many cancers. Several negative regulators accurately quantify the Ras/Raf/ERK pathway, including Ras GTPase-activating phosphatases, proteins, MAPK and the Sprouty/Spred family (21). Since the Sprouty/Spred family is one of the essential suppressors of the Ras/Raf/ERK pathway, many researchers have further investigated the role of Sprouty and Spred as tumor suppressors during tumorigenesis and metastasis (23).

It is believed that Sproutys have several mechanisms to suppress the Ras/Raf/ERK pathway. Sprouty generally acts upstream of Ras because it cannot suppress active Ras mutations (20). However, Sprouty4 inhibits VEGF-A-induced ERK activity by direct binding to c-Raf (Mason et al., 2006). Interestingly, in mammals, Sproutys do not suppress the EGF signaling; Instead, they activate signaling by binding to c-Cbl, an E3 ubiquitin-protein ligase for EGFR (21). Sprouty2 can also suppress Rac1 activation and cell migration through the tyrosine phosphatase 1B (21). In addition, Sprouty1 and Sprouty2 have been reported to be negative regulators of the TGF- β -Smad signaling (24).

Spreds inhibit ERK activation in collaboration with Ras and neurofibromin and suppress phosphorylation and Raf activation (19, 25). Spreds can also regulate the activation of small GTPases, Ras, Rap1, and Rho (19). In Spred1, Ras/Raf/ERK pathway inhibition was known when the two tyrosine residues Y377/Y420 were phosphorylated (26). The other three tyrosine residues, Y303/Y343/Y353 in CRD, are essential for regulating Spred2 activity (27).

The Sprouty/Spred family is induced by many signals from the tyrosine kinase receptor and acts as negative feedback regulators of Ras/Raf/ERK signaling (19-22). The mRNA and protein levels

of members of this family are highly regulated by various mechanisms, including epigenetic and post-translational changes (28, 29).

The Sprouty/Spred family negatively regulates the VEGF-A and VEGF-C signaling pathways, and besides, angiogenesis and lymphangiogenesis are essential processes for tumor development (22). These findings suggest that the expression of the Sprouty/Spred family in the microscopic environment of the tumor also indirectly affects tumorigenesis and metastasis (30, 31).

The Role of Physical Activity, Exercise, and Training on the Ras/Raf/Erk Pathway. However, studies have shown that physical activity increases the expression of ERK genes and proteins (32). In most of these studies, the response of ERK to one or more sessions of activity was evaluated, and immediately after the activity, the level of activity and the amount of phosphorylated ERK was examined. The ERK is activated immediately after the activity in trained and untrained cycling protocols in individuals (33-35), resistance and strength training (36-38), and in situ studies (15, 39, 40). However, few studies have examined the long-term adaptation of this protein and achieved different results. In this regard, it was reported that eccentric contraction of the biceps muscle increases total ERK and phosphorylation after 48 hours, But running downhill after 48 hours did not increase total ERK and phosphorylation (41). Another study reported a significant increase in the total ERK protein content of FHL muscle after eight weeks of resistance training (5 sessions per week) in male Sprague Dawley rats. However, no significant change was observed in its phosphorylated form, as long-term resistance training is probably not a proper intervention to activate ERK (42).

Besides, there are enough reports about the positive effects of various types of physical activity and growth factors. A session of high-intensity interval training (HIIT) on changes in serum vascular endothelial growth factor (VEGF) leads to the onset of the angiogenesis process (43). Also, ten weeks of HIIT, three sessions per week, and 40 minutes per session with an intensity of 70 to 75 maximal heart rate in men with prostate cancer significantly increased VEGF and FGF levels in the experimental group compared to the control group (44).

However, the mechanisms underlying changes in exercise and training growth factors are poorly understood. Since Sprouty/Spred proteins have been shown to act downstream of a wide range of growth factor stimuli, including fibroblast growth factor (FGF), vascular endothelial growth factor (VEGF), platelet growth factor (PDGF), Hepatocyte growth factor, and nerve growth factor (NGF) (45), physical activity, exercise, and training seems to be able to affect Sprouty/Spred by affecting growth factors, which has not yet been studied in this situation.

CONCLUSION

Since most cancers occur in the elderly, it is expected that this disease's incidence and mortality will soon increase. Therefore, paying attention to the cancer control program is necessary globally. Physical activity, exercise, and training are essential health behaviors that play a vital role in the prevention and treatment of cancer, along with different mechanisms to prevent its recurrence and complications of treatment and improve the quality of life of these patients. However, the mechanism of physical activity related to tumor growth inhibition is not fully understood, and the role of new regulators, such as the Sperouty/Spred family, needs to be studied in the future.

ACKNOWLEDGMENT

N/A.

FINANCIAL DISCLOSURE

No financial support was obtained for this study.

FUNDING

This research received no external funding.

ROLE OF THE SPONSOR

There are no sponsors.

ARTIFICIAL INTELLIGENCE (AI) USE

The author declares no AI usage.

CONFLICT OF INTEREST

The authors declare no conflict of interest concerning the authors' contribution and article's publication.

AUTHORS' CONTRIBUTION

Study concept and design: S. M. Tayebi. Acquisition of data: S. M. Tayebi. Analysis and interpretation of data: S. M. Tayebi. Drafting of the manuscript: S. M. Tayebi. Critical revision of the manuscript for important intellectual content: K. Krüger. Statistical analysis: N/A. Administrative, technical, and material support: S. M. Tayebi.

REFERENCES

- 1. Nelson DL, Lehninger AL, Cox MM. Lehninger Principles of Biochemistry. 2th ed: Macmillan Learning; 2012.
- 2. Asada S, Daitoku H, Matsuzaki H, Saito T, Sudo T, Mukai H, et al. Mitogen-activated protein kinases, Erk and p38, phosphorylate and regulate Foxo1. Cellular signalling. 2007;19(3):519-27.
- 3. Keyse SM. An emerging family of dual specificity MAP kinase phosphatases. Biochimica et biophysica acta. 1995;1265(2-3):152-60.
- 4. Boulton TG, Nye SH, Robbins DJ, Ip NY, Radziejewska E, Morgenbesser SD, et al. ERKs: a family of proteinserine/threonine kinases that are activated and tyrosine phosphorylated in response to insulin and NGF. Cell. 1991;65(4):663-75.
- 5. Steelman LS, Chappell WH, Abrams SL, Kempf RC, Long J, Laidler P, et al. Roles of the Raf/MEK/ERK and PI3K/PTEN/Akt/mTOR pathways in controlling growth and sensitivity to therapy-implications for cancer and aging. Aging. 2011;3(3):192-222.
- Casar B, Pinto A, Crespo P. ERK dimers and scaffold proteins: unexpected partners for a forgotten (cytoplasmic) task. Cell cycle (Georgetown, Tex). 2009;8(7):1007-13.
- Sung JH, Kim MO, Koh PO. Nicotinamide prevents the down-regulation of MEK/ERK/p90RSK signaling cascade in brain ischemic injury. The Journal of veterinary medical science. 2012;74(1):35-41.
- Coogan AN, Piggins HD. Circadian and photic regulation of phosphorylation of ERK1/2 and Elk-1 in the suprachiasmatic nuclei of the Syrian hamster. The Journal of neuroscience : the official journal of the Society for Neuroscience. 2003;23(7):3085-93.
- 9. Ma Q-L, Harris-White ME, Ubeda OJ, Simmons M, Beech W, Lim GP, et al. Evidence of Abeta- and transgenedependent defects in ERK-CREB signaling in Alzheimer's models. J Neurochem. 2007;103(4):1594-607.
- 10. Ouwens DM, de Ruiter ND, van der Zon GC, Carter AP, Schouten J, van der Burgt C, et al. Growth factors can activate ATF2 via a two-step mechanism: phosphorylation of Thr71 through the Ras-MEK-ERK pathway and of Thr69 through RalGDS-Src-p38. The EMBO journal. 2002;21(14):3782-93.
- 11. Zhu JH, Gusdon AM, Cimen H, Van Houten B, Koc E, Chu CT. Impaired mitochondrial biogenesis contributes to depletion of functional mitochondria in chronic MPP+ toxicity: dual roles for ERK1/2. Cell death & disease. 2012;3:e312.
- Miyake M, Goodison S, Lawton A, Gomes-Giacoia E, Rosser CJ. Angiogenin promotes tumoral growth and angiogenesis by regulating matrix metallopeptidase-2 expression via the ERK1/2 pathway. Oncogene. 2015;34(7):890-901.
- 13. Wortzel I, Hanoch T, Porat Z, Hausser A, Seger R. Mitotic Golgi translocation of ERK1c is mediated by a PI4KIIIbeta-14-3-3gamma shuttling complex. Journal of cell science. 2015;128(22):4083-95.
- Cheng P, Alberts I, Li X. The role of ERK1/2 in the regulation of proliferation and differentiation of astrocytes in developing brain. International journal of developmental neuroscience : the official journal of the International Society for Developmental Neuroscience. 2013;31(8):783-9.
- 15. Martineau LC, Gardiner PF. Insight into skeletal muscle mechanotransduction: MAPK activation is quantitatively related to tension. Journal of applied physiology (Bethesda, Md : 1985). 2001;91(2):693-702.
- Wretman C, Lionikas A, Widegren U, Lannergren J, Westerblad H, Henriksson J. Effects of concentric and eccentric contractions on phosphorylation of MAPK(erk1/2) and MAPK(p38) in isolated rat skeletal muscle. The Journal of physiology. 2001;535(Pt 1):155-64.
- 17. Roux PP, Blenis J. ERK and p38 MAPK-activated protein kinases: a family of protein kinases with diverse biological functions. Microbiology and molecular biology reviews : MMBR. 2004;68(2):320-44.
- Alavian F, Hajizadeh S, Javan M, Mazloom R. Evaluation Of Erk Activity On Ischemic Tolerance-Induced By Preconditioning With Intermittent Normobaric Hyperoxia In The Rat Model Of Stroke. Arak Medical University Journal. 2017;20(6):41-53 [Article in Farsi].
- Bundschu K, Walter U, Schuh K. Getting a first clue about SPRED functions. BioEssays : news and reviews in molecular, cellular and developmental biology. 2007;29(9):897-907.
- 20. Mason JM, Morrison DJ, Basson MA, Licht JD. Sprouty proteins: multifaceted negative-feedback regulators of receptor tyrosine kinase signaling. Trends in cell biology. 2006;16(1):45-54.
- 21. Masoumi-Moghaddam S, Amini A, Morris DL. The developing story of Sprouty and cancer. Cancer metastasis reviews. 2014;33(2-3):695-720.

- 22. Yoshimura A. Regulation of cytokine signaling by the SOCS and Spred family proteins. The Keio journal of medicine. 2009;58(2):73-83.
- 23. Kawazoe T, Taniguchi K. The Sprouty/Spred family as tumor suppressors: Coming of age. Cancer Sci. 2019;110(5):1525-35.
- 24. Shin EH, Basson MA, Robinson ML, McAvoy JW, Lovicu FJ. Sprouty is a negative regulator of transforming growth factor beta-induced epithelial-to-mesenchymal transition and cataract. Molecular medicine (Cambridge, Mass). 2012;18:861-73.
- 25. Stowe IB, Mercado EL, Stowe TR, Bell EL, Oses-Prieto JA, Hernandez H, et al. A shared molecular mechanism underlies the human rasopathies Legius syndrome and Neurofibromatosis-1. Genes & development. 2012;26(13):1421-6.
- 26. Quintanar-Audelo M, Yusoff P, Sinniah S, Chandramouli S, Guy GR. Sprouty-related Ena/vasodilator-stimulated phosphoprotein homology 1-domain-containing protein (SPRED1), a tyrosine-protein phosphatase non-receptor type 11 (SHP2) substrate in the Ras/extracellular signal-regulated kinase (ERK) pathway. The Journal of biological chemistry. 2011;286(26):23102-12.
- 27. Meng S, Zhang M, Pan W, Li Z, Anderson DH, Zhang S, et al. Tyrosines 303/343/353 within the Sprouty-related domain of Spred2 are essential for its interaction with p85 and inhibitory effect on Ras/ERK activation. The international journal of biochemistry & cell biology. 2012;44(5):748-58.
- 28. Gao X, Hicks KC, Neumann P, Patel TB. Hypoxia inducible factors regulate the transcription of the sprouty2 gene and expression of the sprouty2 protein. PLOS ONE. 2017;12(2):e0171616.
- 29. Sun J, Zhang J, Wang Y, Li Y, Zhang R. A Pilot Study of Aberrant CpG Island Hypermethylation of SPRED1 in Acute Myeloloid Leukemia. International journal of medical sciences. 2019;16(2):324-30.
- 30. Taniguchi K, Ishizaki T, Ayada T, Sugiyama Y, Wakabayashi Y, Sekiya T, et al. Sprouty4 deficiency potentiates Ras-independent angiogenic signals and tumor growth. Cancer Sci. 2009;100(9):1648-54.
- 31. Taniguchi K, Sasaki K-i, Watari K, Yasukawa H, Imaizumi T, Ayada T, et al. Suppression of Sproutys Has a Therapeutic Effect for a Mouse Model of Ischemia by Enhancing Angiogenesis. PLOS ONE. 2009;4(5):e5467.
- 32. Kim Y, Inoue T, Nakajima R, Nakae K, Tamura T, Tokuyama K, et al. Effects of endurance training on gene expression of insulin signal transduction pathway. Biochemical and biophysical research communications. 1995;210(3):766-73.
- 33. Widegren U, Jiang XJ, Krook A, Chibalin AV, Bjornholm M, Tally M, et al. Divergent effects of exercise on metabolic and mitogenic signaling pathways in human skeletal muscle. FASEB journal : official publication of the Federation of American Societies for Experimental Biology. 1998;12(13):1379-89.
- 34. Widegren U, Wretman C, Lionikas A, Hedin G, Henriksson J. Influence of exercise intensity on ERK/MAP kinase signalling in human skeletal muscle. Pflugers Archiv : European journal of physiology. 2000;441(2-3):317-22.
- 35. Yu M, Stepto NK, Chibalin AV, Fryer LG, Carling D, Krook A, et al. Metabolic and mitogenic signal transduction in human skeletal muscle after intense cycling exercise. The Journal of physiology. 2003;546(Pt 2):327-35.
- 36. Creer A, Gallagher P, Slivka D, Jemiolo B, Fink W, Trappe S. Influence of muscle glycogen availability on ERK1/2 and Akt signaling after resistance exercise in human skeletal muscle. Journal of applied physiology (Bethesda, Md : 1985). 2005;99(3):950-6.
- 37. Karlsson HK, Nilsson PA, Nilsson J, Chibalin AV, Zierath JR, Blomstrand E. Branched-chain amino acids increase p70S6k phosphorylation in human skeletal muscle after resistance exercise. American journal of physiology Endocrinology and metabolism. 2004;287(1):E1-7.
- 38. Williamson D, Gallagher P, Harber M, Hollon C, Trappe S. Mitogen-activated protein kinase (MAPK) pathway activation: effects of age and acute exercise on human skeletal muscle. The Journal of physiology. 2003;547(Pt 3):977-87.
- 39. Nader GA, Esser KA. Intracellular signaling specificity in skeletal muscle in response to different modes of exercise. Journal of applied physiology (Bethesda, Md : 1985). 2001;90(5):1936-42.
- 40. Sherwood DJ, Dufresne SD, Markuns JF, Cheatham B, Moller DE, Aronson D, et al. Differential regulation of MAP kinase, p70(S6K), and Akt by contraction and insulin in rat skeletal muscle. The American journal of physiology. 1999;276(5):E870-8.
- 41. Thompson HS, Maynard EB, Morales ER, Scordilis SP. Exercise-induced HSP27, HSP70 and MAPK responses in human skeletal muscle. Acta physiologica Scandinavica. 2003;178(1):61-72.
- 42. Nemati J, Samadi M, Hadidi V, Ghodrat L. The effect of 8 weeks of resistance training on total and phosphorylated extracellular signal regulated kinases (ERK) in flexor hallucis longusmuscle of rats. Journal of Practical Studies of Biosciences in Sport (JPSBS). 2018;6(12):117-26 [Article in Farsi].
- 43. Bayati M, Gharakhanlou R, Nikkhah M, Amani Shalamzari S. The Effect of Four Weeks of High-intensity Interval Training on PGC-1α and VEGF Protein Contents in Skeletal Muscle of Active Men. Journal of Arak University of Medical Science. 2018;21(3):24-32 [Article in Farsi].

- 44. Fathollahi F, Faramarzi M, Hemmati R. The Effect of 10 weeks of high-intensity exercise training on resting levels of some angiogenesis and pulmonary function of men with prostate cancer. Journal of Fasa University of Medical Science. 2019;8(4):1097-105 [Article in Farsi].
- 45. Guy GR, Jackson RA, Yusoff P, Chow SY. Sprouty proteins: modified modulators, matchmakers or missing links? The Journal of endocrinology. 2009;203(2):191-202.